

# Package: grmsem (via r-universe)

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**Title** Genetic-Relationship-Matrix Structural Equation Modelling (GRMSEM)

**Version** 1.1.0

**Maintainer** Beate StPourcain <Beate.StPourcain@mpi.nl>

**Description** Quantitative genetics tool supporting the modelling of multivariate genetic variance structures in quantitative data. It allows fitting different models through multivariate genetic-relationship-matrix (GRM) structural equation modelling (SEM) in unrelated individuals, using a maximum likelihood approach. Specifically, it combines genome-wide genotyping information, as captured by GRMs, with twin-research-based SEM techniques, St Pourcain et al. (2017) <[doi:10.1016/j.biopsych.2017.09.020](https://doi.org/10.1016/j.biopsych.2017.09.020)>, Shapland et al. (2020) <[doi:10.1101/2020.08.14.251199](https://doi.org/10.1101/2020.08.14.251199)>.

**Depends** R (>= 3.5)

**Imports** msm (>= 1.6), numDeriv, optimParallel, stats, utils

**License** GPL-3

**Encoding** UTF-8

**LazyData** true

**RoxygenNote** 7.1.0

**Suggests** bookdown, knitr, rmarkdown, testthat

**VignetteBuilder** knitr

**NeedsCompilation** no

**URL** <https://CRAN.R-project.org/package=grmsem>,  
<https://gitlab.gwdg.de/beate.stpourcain/grmsem>

**BugReports** <https://gitlab.gwdg.de/beate.stpourcain/grmsem/-/issues>

**Author** Beate StPourcain [aut, cre]  
(<<https://orcid.org/0000-0002-4680-3517>>), Alexander Klassmann [ctb]

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fit.large

*Prefitted model: large data set*

---

## Description

A large quad-variate data set was simulated assuming an underlying Cholesky model, with 5000 observations per trait and high polygenicity (20,000 SNPs per genetic factor). Genetic trait variances were set to 0.30, 0.60, 0.60 and 0.70 respectively, and residual variances to 0.70, 0.40, 0.40 and 0.30. The data set is described in full, including genetic and residual covariances, in the vignette. A 4-variate Cholesky model was fitted to the data as described in the vignette and the output has been saved in fit.large.RData.

## Usage

fit.large

## Format

gsem.fit output (a list object)

**model.in** input parameters

**formula** fitted formula

**model.fit** optimisation output

**model.out** fitted gsem model

**VCOV** variance covariance matrix

**k** Number of phenotypes  
**n** Number of observations across all phenotypes  
**n.obs** Number of observations per phenotype  
**n.ind** Number of individuals with at least one phenotype  
**model** gsem model  
**con** constraint  
**ph.nms** phenotype names

---

G.small

*Symmetric GRM data: small data set*


---

### Description

A genetic relationship matrix for a small tri-variate data set was simulated assuming an underlying Cholesky model, with 100 observations per trait and low polygenicity (150 SNPs per genetic factor). Genetic trait variances were set to 0.30, 0.60, 0.60 and 0.70 respectively, and residual variances to 0.70, 0.40, 0.40 and 0.30. The data set is described in full, including genetic and residual covariances, in the vignette. The traits are influenced by three independent genetic factors (A1, A2 and A3, based on a 150 SNPs each) and three independent residual factors (E1, E2 and E3).

### Usage

G.small

### Format

Symmetric GRM data frame with 100 rows and 100 columns (observations)

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grm.bin.input

*grm import function*


---

### Description

This function imports genetic relationship matrices (GRMs) in binary format as e.g. stored by the GCTA software command `gcta64 --grm test --make-grm-bin --out test`

### Usage

```
grm.bin.input(file, size = 4)
```

### Arguments

**file** name of the binary grm file. No default.  
**size** byte size used (typically gcta uses 4). See eponymous parameter of function [readBin](#).

**Value**

object of type matrix

grmse**m**.bin.input imports a binary GCTA GRM (.bin) file and transforms it into a symmetric matrix

---

gr <b>m</b> .input	<i>gr<b>m</b> import function</i>
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**Description**

This function imports genetic relationship matrices (GRMs) in gz format as e.g. stored by the GCTA software command `gcta64 --grm test --make-grm-gz --out test`

**Usage**

```
grm.input(file)
```

**Arguments**

file                    the name of the gz compressed gr**m** file. No default.

**Value**

grmse**m**.input imports a zipped GCTA GRM (.gz) file and transforms it into a symmetric matrix

---

grmse <b>m</b> .biher	<i>grmse<b>m</b> bivariate heritability estimation function.</i>
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---

**Description**

This function estimates the bivariate heritability.

**Usage**

```
grmsem.biher(ph, grmsem.var.out = NULL)
```

**Arguments**

ph                      Phenotype file as R dataframe (columns:  $\geq 2$  phenotypes, rows:  $n_i$  individuals in the same order as G). No default.

grmse**m**.var.out      A grmse**m**.var object with unstandardised parameters (factor loadings). Default NULL.

**Details**

The `grmsem.biher` function estimates the bivariate heritability (DS, Cholesky, IP and IPC models) from the observed phenotype data and a `grmsem.var` object. All standard errors are derived with the Delta method.

**Value**

`grmsem.biher` returns a list object consisting of the following matrices:

VPO	observed phenotypic variance/covariance matrix
VA	estimated genetic variance
BIHER	estimated bivariate heritability (off-diagonals): VA / VPO
BIHER.se	standard error of estimated bivariate heritability
BIHER.Z	Z (wald) of estimated bivariate heritability
BIHER.p	p (Wald) of estimated bivariate heritability

**Examples**

```

#(runtime should be less than one minute)

out <- grmsem.fit(ph.small, G.small, LogL = TRUE, estSE = TRUE)
var.out <- grmsem.var(out)
grmsem.biher(ph.small, var.out)

```

---

<code>grmsem.fcoher</code>	<i>grmsem factorial co-heritability and co-environmentality estimation function.</i>
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**Description**

This function estimates factorial co-heritabilities and factorial co-environmentalities.

**Usage**

```
grmsem.fcoher(grmsem.out = NULL)
```

**Arguments**

`grmsem.out` `grmsem.fit` or `grmsem.stpar` object. Default NULL.

**Details**

The `grmsem.fcoher` function can be used to estimate factorial co-heritabilities and factorial co-environmentalities for models estimating latent variables (Cholesky, IP or IPC models), based on `grmsem.fit` or `grmsem.stpar` objects. The factorial co-heritability of a genetic factor  $m$  for trait  $t$  is the ratio of the genetic variance explained by factor  $m$  ( $A_{mt}$ ) to the total genetic variance ( $A_t$ ):  $A_{mt} / A_t$ . The factorial co-environmentality of a residual factor  $n$  for trait  $t$  is the ratio of the residual variance explained by factor  $n$  ( $E_{nt}$ ) to the total residual variance ( $E_t$ ):  $E_{nt} / E_t$ . All standard errors are derived with the Delta method.

**Value**

grmsem.fcoher returns an extended model.out dataframe, fcoher.model.out, with the following columns:

label	parameter label
estimates	estimated parameters
gradient	gradient
se	SE
Z	Z (Wald) of factor loading
p	p (Wald) of factor loading
Vi	squared factor loading, explained phenotypic variation by the factor
Vi.se	SE of squared factor loading
FCOHER	factorial co-heritability
FCOHER.se	SE of factorial co-heritability
FCOHER.Z	Z (wald) of factorial co-heritability
FCOHER.se	p (Wald) of factorial co-heritability
FCOENV	factorial co-environmentality
FCOENV.se	SE of factorial co-environmentality
FCOENV.Z	Z (wald) of factorial co-environmentality
FCOENV.se	p (Wald) of factorial co-environmentality

**Examples**

```

#(runtime should be less than one minute)

out <- grmsem.fit(ph.small, G.small, LogL = TRUE, estSE = TRUE)
grmsem.fcoher(out)

```

---

grmsem.fit

*grmsem model fitting function*


---

**Description**

This function fits a grmsem model.

**Usage**

```
grmsemlfit(
  ph,
  G,
  A.v.free = NULL,
  E.v.free = NULL,
  A.v.startval = NULL,
  E.v.startval = NULL,
  LogL = FALSE,
  estSE = FALSE,
  cores = 1,
  model = "Cholesky",
  compl.ph = FALSE,
  printest = FALSE,
  cluster = "PSOCK",
  optim = "optim",
  verbose = FALSE
)
```

**Arguments**

ph	phenotype file as R dataframe, even for single vectors (columns: k phenotypes, rows: ni individuals in same order as G). No default.
G	GRM matrix as provided by the grm.input or grm.bin.input.R function. Use the same order of individuals as in ph. No default.
A.v.free	vector of free parameters for genetic factor loadings (free:1, not-free:0). Default NULL, all parameters are estimated.
E.v.free	vector of free parameters for residual factor loadings (free:1, not-free:0). Default NULL, all parameters are estimated.
A.v.startval	vector of starting values for genetic factor loadings. Default NULL, all starting values are generated.
E.v.startval	vector of starting values for residual factor loadings. Default NULL, all starting values are generated.
LogL	estimation of the loglikelihood using optim BFGS (TRUE/FALSE). Default FALSE.
estSE	estimation of standard errors by recalculating the Hessian matrix. Default FALSE.
cores	number of cores for multi-threaded calculations (numeric). Default 1.
model	grmseml model selection. Options: "Cholesky", "IP", "IPC", "DS". Default "Cholesky".
compl.ph	listwise complete observations across all phenotypes (all NA are excluded). Default FALSE.
printest	print output of the model.fit function including estimates (printest.txt) that can be used as starting values. Default FALSE.
cluster	cluster type. Options: "PSOCK", "FORK". Default "PSOCK".

optim	optimisation function from stats or optimParallel libraries. Options: "optim", "optimParallel". Default "optim".
verbose	additional model fit information: (i) phenotype vector, (ii) n of GRM and corresponding I matrix when data are missing, (iii) Hessian if estSE TRUE. Default FALSE.

## Details

grmsem models estimate genetic (A) and residual (E) variance/covariance of quantitative traits (AE model), where E in GRM-based methods can capture both, shared and unique residual influences. The estimation of parameters and their SEs is performed with the function `grmsem.fit`. Specifically, the loglikelihood is estimated with `stats::optim` and the BFGS (Broyden-Fletcher-Goldfarb-Shannon) approach and the variance/covariance matrix of estimated parameters with `numDeriv::hessian`. The statistical significance of estimated parameters is assessed using a Wald test, assuming multivariate normality.

`grmsem.fit` allows fitting different models describing the underlying multivariate genetic architecture of quantitative traits, as captured by a genetic-relationship-matrix (GRM), using structural equation modelling (SEM) techniques and a maximum likelihood approach. The user can fit multiple predefined model structures to the data. A Cholesky decomposition, Independent Pathway, and hybrid Independent Pathway/Cholesky models can be fitted by setting the `model` option to Cholesky, IP or IPC, respectively. In addition, the Cholesky model can be re-parametrised as Direct Symmetric model, estimating genetic and residual covariances directly, using the `model` option DS. Each model can be adapted by the user by setting free parameters (`A.v.free` and `E.v.free` options) and starting values (`A.v.startval` and `E.v.startval` options).

Input parameters are returned as `model.in` list object. Output from the maximum likelihood estimation is also given as list `model.fit` and the fitted grmsem model with estimated parameters and SEs is returned as dataframe `model.out`. The returned `grmsem.fit` object can be used to estimate genetic and residual covariance and correlations (`grmsem.var` function), bivariate heritabilities (`grmsem.biher` function), and factorial co-heritabilities and co-environmentalities (`grmsem.fcoher` function). All estimated parameters of the fitted grmsem model can also be standardised using the function `grmsem.stpar`.

Listwise complete observations can be selected with the option `compl.ph=TRUE`. Otherwise, `grmsem.fit` fits, like GREML, all available data to the model with the default option `compl.ph FALSE`. Using the option `LogL=FALSE`, the user can check the model input parameters and formula without a maximum likelihood estimation. Using the option `estSE=FALSE`, the user can carry out a maximum likelihood estimation without the estimation of SEs for estimated parameters that require calculating the Hessian. Note that `grmsem.fit` should preferably be run in parallel, by setting the `cores` option to the required number of cores.

When `grmsem.fit` is called with `LogL TRUE`, the user will see also the iterations of the `stats::optim` loglikelihood estimation, which are not included in the exported `grmsem.fit` list object.

## Value

`grmsem.fit` returns a `grmsem.fit` list object consisting of:

<code>model.in</code>	list of input parameters
<code>formula</code>	matrix of the model specification



model.fit list output of the maximum likelihood estimation, if LogL TRUE  
 model.out dataframe of fitted grmsem model with estimated parameters and SEs, if estSE TRUE  
 VCOV variance/covariance matrix  
 k number of phenotypes  
 n total number of observations across all phenotypes  
 n.obs number of observations per phenotype  
 n.ind number of individuals with at least one phenotype  
 model type of grmsem model  
 ph.nms vector of phenotype names

model.in list of input parameters:

part a - genetic, e - residual parameters  
 label parameter label  
 value starting values  
 freepar free parameters

model.fit list output of the maximum likelihood estimation:

optimisation output via optim()  
 estimates estimated parameters: factor loadings for 'Cholesky', 'IP' and 'IPC' models, but variance components for 'DS'  
 LL loglikelihood  
 calls optim() calls  
 convergence optim() convergence  
 message optim() message

model.out data.frame of fitted grmsem model:

label parameter label  
 estimates estimated parameters  
 gradient gradient  
 se SE  
 Z Z (Wald)  
 p p (Wald)

## Examples

```

#Set up a Cholesky model: Model formula and total number of parameters
#ph.small is a trivariate phenotype file for 100 individuals in the same order as G.small
#nrow = 100, ncol = 3
#(runtime should be less than one minute)
out <- grmsem.fit(ph.small, G.small, LogL = FALSE, estSE = FALSE)

#Run a Cholesky model:
out <- grmsem.fit(ph.small, G.small, LogL = TRUE, estSE = TRUE)

```

---

grmsem.stpar	<i>grmsem standardised parameters function</i>
--------------	--

---

**Description**

This function estimates standardised parameters for a grmsem.fit object.

**Usage**

```
grmsem.stpar(grmsem.out = NULL)
```

**Arguments**

grmsem.out      grmsem.fit object as provided by the grmsem.fit function. Default NULL.

**Details**

grmsem.stpar standardises grmsem.fit estimates so that derived or estimated A and E variance components will add up to phenotypic unit variance. The SEs of standardised parameters are derived using the Jacobian matrix.

**Value**

grmsem.stpar returns a list object consisting of:

model.in	list of input parameters
stand.model.out	dataframe of fitted grmsem model with standardised parameters and SEs
stVCOV	standardised variance/covariance matrix
k	number of phenotypes
n	total number of observations across all phenotypes
model	type of grmsem model
ph.nms	vector of phenotype names

model.in list of input parameters:

part	a - genetic, e - residual parameters
label	parameter label
value	starting values
freepar	free parameters

stand.model.out dataframe of fitted grmsem model with standardised parameters and SEs:

label	parameter label
estimates	standardised estimated parameters
se	SE
Z	Z (Wald)
p	p (Wald)

**Examples**

```

#(runtime should be less than one minute)

out <- grmseml.fit(ph.small, G.small, LogL = TRUE, estSE = TRUE)
stout <- grmseml.stpar(out)
print(stout)

```

---

grmseml.var	<i>grmseml variance estimation function</i>
-------------	---

---

**Description**

This function estimates genetic and residual variances, and genetic correlations.

**Usage**

```
grmseml.var(grmseml.out = NULL)
```

**Arguments**

grmseml.out      A grmseml.fit or grmseml.stpar object. Default NULL.

**Details**

The grmseml.var function can be used to estimate genetic and residual covariance and correlations for DS, Cholesky, IP and IPC models, based on grmseml.fit or grmseml.stpar objects. For the latter, the diagonal elements of the VA output matrix detail the heritabilities. Except for directly estimated variance components using the DS model, all standard errors are derived with the Delta method.

**Value**

grmseml.var returns a list object consisting of the following matrices:

VA	estimated genetic variance
VA.se	standard error of estimated genetic variance
VE	estimated residual variance
VE.se	standard error of estimated residual variance
VP	estimated total phenotypic variance
RG	genetic correlation
RG.se	standard error genetic correlation
RE	residual correlation
RG.se	standard error residual correlation

**Examples**

```
 #(runtime should be less than one minute)

 out <- grmsem.fit(ph.small, G.small, LogL = TRUE, estSE = TRUE)
 var.out <- grmsem.var(out)
 print(var.out)
```

---

ph.large                      *Phenotype data: large data set*

---

**Description**

A large quad-variate data set was simulated assuming an underlying Cholesky model, with 5000 observations per trait and high polygenicity (5,000 SNPs per genetic factor). The data set is described in full in the vignette. Genetic and residual covariances are assumed to be influenced by four independent genetic factors (A1, A2, A3 and A4, based on 5000 SNPs each) and four independent residual factors (E1, E2, E3 and E4), respectively.

**Usage**

```
ph.large
```

**Format**

A data frame with 5000 rows and 4 columns

**Y1** trait one

**Y2** trait two

**Y3** trait three

**Y4** trait four

---

ph.small                      *Phenotype data: small data set*

---

**Description**

A small tri-variate data set was simulated assuming an underlying Cholesky model, with 100 observations per trait and low polygenicity (150 SNPs per genetic factor). The data set is described in full in the vignette. Genetic and residual covariances are assumed to be influenced by three independent genetic factors (A1, A2 and A3, based on a 150 SNPs each) and three independent residual factors (E1, E2 and E3), respectively.

**Usage**

```
ph.small
```

**Format**

A data frame with 100 rows and 3 columns

**Y1** trait one

**Y2** trait two

**Y3** trait three

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